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**Parenteral and Ophthalmic Drug Products Leachables and Extractables Working Group**

**Issued and Effective  
September, 2011**

Study Protocol – Stage 1  
Amendment #1

*Experimental Protocol for Qualitative Controlled Extraction Studies on Material Test Articles Representative of Prefilled Syringe (PFS) and Small Volume Parenteral (SVP) Container Closure Systems*

**TABLE OF CONTENTS**

26		
27		
28	<b>I. INTRODUCTION; PURPOSE OF AMENDMENT #1</b>	<b>2</b>
29		
30	<b>II. PURPOSE AND SCOPE OF WORK (STUDY PROTOCOL STAGE 1)</b>	<b>2</b>
31		
32	<b>III. REGULATORY STATUS</b>	<b>3</b>
33		
34	<b>IV. SAFETY AND ENVIRONMENTAL IMPACT</b>	<b>3</b>
35		
36	<b>V. TEST ARTICLES</b>	<b>4</b>
37		
38	<b>VI. CHEMICALS AND EQUIPMENT</b>	<b>4</b>
39		
40	A. Extraction Solvents and Additional Chemicals	4
41	B. Extraction Equipment	5
42	C. Analytical Instrumentation	5
43		
44	<b>VII. EXTRACTION PROCEDURES</b>	<b>6</b>
45		
46	A. General	6
47	B. Extraction Maps	7
48	C. General Considerations	8
49	D. Soxhlet Extraction	9
50	E. Reflux	9
51	F. Sealed Vessel Extraction	9
52		
53	<b>VIII. ANALYTICAL METHODS</b>	<b>9</b>
54		
55	A. General	9
56	B. Gas Chromatography (GC)	10
57	C. High Performance Liquid Chromatography (HPLC)	10
58	D. Inductively Coupled Plasma Atomic Spectroscopy (ICPAS)	10
59	E. Headspace GC/MS Analysis	10
60		
61	<b>IX. DATA EVALUATION AND REPORTING</b>	<b>10</b>
62		
63	A. Qualitative Analysis	10
64	B. Semi-Quantitative Measurement	11
65		
66	<b>X. GLOSSARY</b>	<b>11</b>
67		
68	<b>XI. REFERENCES</b>	<b>12</b>
69		

70 **I. Introduction; Purpose of Amendment #1**  
71

72 The original Protocol for this study included five Test Articles, as specified in Table I of that  
73 document. Three additional Test Articles, a label and a low-density polyethylene bottle and its  
74 associated polypropylene cap, are added to this study as such Articles may be relevant to the  
75 types of packaging systems utilized with PODP drug products.  
76

77 **II. Purpose and Scope of Work (Study Protocol Stage I)**  
78

79 The purpose of the experiments outlined in this protocol is to generate data from Controlled  
80 Extraction Studies, which the Working Group will use to investigate its hypotheses:  
81

- 82 1. Threshold concepts that have been developed for safety qualification of leachables in  
83 OINDP can be extrapolated to the evaluation and safety qualification of leachables in  
84 PODP, with consideration of factors and parameters such as dose, duration, patient  
85 population and product dependent characteristics unique to various PODP types.  
86
- 87 2. The science-based best demonstrated practices established for the OINDP pharmaceutical  
88 development process can be extrapolated to PODP container closure systems.  
89
- 90 3. Threshold and best practices concepts can be integrated into a comprehensive process for  
91 characterizing container closure systems with respect to leachable substances and their  
92 associated impact on PODP safety.  
93

94 Controlled Extraction Studies will be performed following the general methodologies contained  
95 in this protocol. Test articles will be subjected to different extraction conditions to establish how  
96 different experimentally controlled parameters affect the resulting extractables profiles. Of  
97 specific interest to the Working Group are the parenteral and ophthalmic dosage forms,  
98 particularly Small Volume Parenterals (SVP), Large Volume Parenterals (LVP), Pre-filled  
99 Syringes (PFS) and Blow-Fill-Seal systems (BFS). The Stage 1 Protocol specifically focuses on  
100 the SVP and PFS dosage forms and on the generation of qualitative extractables profiles. Future  
101 Stages will focus on additional dosage forms and/or quantitative aspects of extractables profiling.  
102 The intent of the Stage 1 assessment is to generate the fundamental information from which Best  
103 Demonstrated Practices can be derived; it is not the intent of this Stage 1 assessment to  
104 prospectively establish the practices used in this study as the Best Demonstrated Practices  
105 themselves.  
106

107 As no single analytical technique can be used to identify and quantify all unknown extractables,  
108 a variety of methods will be utilized in this protocol to maximize the likelihood that all  
109 predominant extractable compounds associated with the test articles are accounted for and  
110 appropriately evaluated. Overlap between methods will supply corroborating data that  
111 demonstrate the validity of the procedures. To provide a full analytical survey of possible  
112 analytes the following strategy will be employed:  
113

- 114 1. Gas Chromatography with appropriate sampling/injection and detection strategies  
115 e.g. Flame Ionization Detection (GC/FID) and Mass Spectrometry (GC/MS)] for  
116 identification and assessment of volatile and semi-volatile extractables.  
117 2. High Performance Liquid Chromatography with appropriate detection strategies  
118 [e.g. Diode Array Detection (HPLC/DAD), Mass Spectrometry (LC/MS)] for  
119 identification and assessment of relatively polar and non-volatile extractables.  
120 3. Inductively Coupled Plasma/Mass Spectrometry (ICP/MS) and/or Inductively  
121 Coupled Plasma/Atomic Emission Spectroscopy (ICP/AES) to detect single  
122 elements in the extracts (i.e. metals).  
123

124 While analytical tests and measurements, such as pH, UV absorbance, and total organic carbon  
125 (TOC), can provide insight into the general chemical nature and amount of extracted substances,  
126 they do not directly provide information for the identification and/or quantitation of individual  
127 extractables and thus will not be utilized in this study.  
128

129 Studies designed to assess recovery (i.e. mass balance) for individual extractables relative to the  
130 known formulations of chemical additives in the various test articles, or reproducibility of  
131 extractables profiles for multiple “batches” of any particular test article are not within the scope  
132 of this Stage of the test protocol. Additionally, the extraction procedures, analytical  
133 techniques/methods, and analysis conditions described in this experimental test protocol will not  
134 be fully and rigorously validated. Nevertheless, the scientific credibility of the data generated in  
135 this study shall be established via the utilization of system suitability testing with all the analysis  
136 methods and by the expert review of the generated data. Finally, “special case” classes of  
137 extractables that have defined and highly specific analytical methods that are generally accepted  
138 and commonly used for their identification and quantitative assessment will not be considered in  
139 this study.  
140

### 141 **III. REGULATORY STATUS**

142

143 This experimental test protocol will be conducted in the spirit of Good Laboratory Practices and  
144 Good Manufacturing Practices (GXP) requirements. All experiments shall be documented based  
145 on the appropriate GXP compliance systems in a participating laboratory. Any changes or  
146 clarifications that a participating laboratory makes to this test protocol shall be documented as  
147 appropriate, and discussed/approved by the Study Coordination as appropriate.  
148

### 149 **IV. SAFETY AND ENVIRONMENTAL IMPACT**

150

151 Chemicals and reagents used in this study (e.g. organic solvents commonly used to enhance  
152 solubility of lipophilic targets and to increase transport of small molecules out of complex  
153 matrices) may be flammable and/or pose short-term and long-term environmental health risks.  
154 Care must be exercised with their use. Consult the Material Safety and Data Sheet (MSDS) for  
155 appropriate personal protection and disposal. Safety risks associated with the various processes  
156 and procedures performed in this study may exist and should be understood and managed using  
157 such strategies as environmental control and personal protection.  
158

159 **V. TEST ARTICLES**

160  
 161 A list of the test articles available for use in this study is provided in Table 1. Test articles will  
 162 be provided in an appropriate form for use as test articles. Certain, but not necessarily all, details  
 163 of the additive formulations and manufacturing conditions for these test articles are known and  
 164 are captured in Table 1.  
 165

TABLE 1. TEST ARTICLES.			
MATERIAL TYPE	MATERIAL APPLICATION	MATERIAL FORMAT	COMPOSITION
Low density polyethylene (LDPE)	Bottle/ Vial	Bottle	4 oz LDPE, part B347A (Container & Packaging Supply)
Polypropylene (PP)	Cap	Cap	PP, Part L764(Container & Packaging Supply)
Adhesive Label	Label applied to outside of PODP Packaging Systems	Sheets	<p><u>Substrate:</u> Unknown</p> <p><u>Adhesive:</u> Acrylic polymer(s), residual monomers, water, ammonia (99.55%); wetting agent, Surfynol 336, at 0.4% containing CAS 577-11-7 (&gt; 25%), CAS 9014-85-1 (&gt; 25%); Biocide, Kathon LX, at 0.05% containing Chloro-2-methyl-4-isothiazolin-3-one (CAS 26172-55-4), 1.1-1.4%, 2-Methyl-4-isothiazolin-3-one (CAS 2682-20-4), 0.3 - 0.5%, Magnesium Chloride (CAS 7786-30-3), 1.0 - 1.2%, Magnesium nitrate (CAS 10377-60-3), 1.4 - 2.0% Copper nitrate (CAS 3251-23-8) 1,500 - 1,700 ppm, Water, 95 - 97%</p> <p><u>Printing ink:</u> Irgacure 369 (CAS 119313-12-1) and Irgacure 1173 (CAS 7473-98-5), photoinitiators; Trimethylolpropane triacrylate (TMPTA, CAS 15625-89-5), Tripropylene glycol diacrylate (TPGDA, CAS 42978-66-5), Glycerol propoxy triacrylate (GPTA, CAS 52408-84-1), monomers; HQME/Mequinol (CAS 150-76-5), stabilizer; Carbon black (CAS 1333-86-4), Phthalo blue (CAS 147-14-8), Carbazole violet (CAS 215247-95-3), pigments</p> <p><u>Varnish:</u> Unknown</p>

166  
 167  
 168 **VI. CHEMICALS AND EQUIPMENT**

169  
 170 Extraction and analytical methods were chosen and designed to utilize chemicals, apparatus, and  
 171 instrumentation available in typical laboratories routinely involved with this type of study.

172  
 173 **A. Extraction Solvents and Additional Chemicals**

174  
 175 The chemicals required for use as, or in preparation of, extraction solvents, as well as the  
 176 directions for the preparation of several of these extraction solvents, were outlined in the original  
 177 Protocol and such information is directly relevant to the testing to be performed as a result of this Protocol  
 178 Amendment #1.  
 179  
 180

181 **B. Extraction Equipment**

- 182
- 183 1. Soxhlet Extraction
- 184 • Soxhlet apparatus.
- 185 • All glass labware for these extractions must be acid-washed prior to use.
- 186 • The use of any lubricants, such as vacuum grease on ground glass joints, should be
- 187 avoided.
- 188
- 189 2. Reflux
- 190 • Reflux apparatus [e.g. round bottom flask (200 mL or larger), condenser with ground
- 191 glass joints, hot plate or heating mantle].
- 192 • All glass labware for these extractions must be acid-washed prior to use.
- 193 • The use of any lubricants, such as vacuum grease on ground glass joints, should be
- 194 avoided.
- 195 3. Sealed Container
- 196 • Teflon [Savillex (6133 Baker Road, Minnetonka, MN 55345-5910 USA, Phone: 952-
- 197 935-4100, E-mail: [info@savillex.com](mailto:info@savillex.com)), Part # 0108, 8 fl. Oz. Teflon Jar]
- 198 • Pyrex [VWR (Customer Service: 1-800-932-5000), Catalog # 89000-236, Media /
- 199 Storage Bottles with Standard GL45 Polypropylene Cap, 250 mL] containers
- 200 • All glass labware for these extractions must be acid-washed prior to use. Teflon
- 201 vessels are used with the high pH extractions to avoid any leaching from glass,
- 202 especially for samples for ICP analysis
- 203 • Autoclave
- 204 • Oven with operating range of 30 to 75 °C; explosion proof
- 205

206 **C. Analytical Instrumentation**

- 207
- 208 • Gas chromatograph equipped with a Flame Ionization Detector (GC/FID)
- 209 • Gas chromatograph equipped with a Mass Spectrometer (GC/MS). GC systems that employ
- 210 flow splitting to accomplish FID and MS detection in tandem could be used in this study.
- 211 • Headspace Sampler/Injector (HS) for GC/MS Instrumentation.
- 212 • Liquid chromatograph equipped with a photodiode array detector
- 213 • Liquid chromatograph equipped with an APCI (Atmospheric Pressure Chemical Ionization)
- 214 capable Mass Spectrometer (LC/MS). Preference is given to LC systems that are capable of
- 215 both DAD and MS detection. Additional detectors (e.g. corona assisted discharge detectors,
- 216 evaporative light scattering) may be used as appropriate.
- 217 • Inductively Coupled Plasma Mass Spectrometer (ICP-MS)
- 218
- 219
- 220
- 221

222 **VII. EXTRACTION PROCEDURES**

223

224 **A. General**

225

226 In the PQRI OINDP studies, extractions were performed on each test article using three solvents  
227 representing a range of polarity, specifically

228

229 • methylene chloride (dichloromethane)

230 • 2-propanol (isopropanol, IPA)

231 • hexane (n-hexane, not hexanes).

232

233 This was appropriate in the case of OINDP given the nature of the drug vehicles used in those  
234 types of products (organic solvents) and the conditions of contact between the drug vehicles and  
235 the container closure system (continuous direct contact over shelf life).

236

237 While the use of such extraction solvents may be relevant for PODP products, a significant  
238 portion of PODP products are water-based and the three solvents previously employed do not  
239 address the unique solubilizing properties of water and aqueous buffer systems. Thus in the case  
240 of PODP, the OINDP solvents will be augmented by aqueous extraction media. These additional  
241 aqueous extraction media, and their associated justification, include

242

243 \* Water at pH 2.5 (HCl/KCl mixture); justification, few therapeutic products are lower than pH  
244 2.5.

245 \* Water at pH 9.5 (Phosphate buffer); justification, few therapeutic products are higher in pH  
246 than 9.5.

247 \* 1/1 IPA/water; justification; simulates aqueous formulations containing solubilizing agents,  
248 provides for trend analysis (with IPA and water alone).

249

250 Thus, the five extraction media to be used in this Stage 1 Protocol are the three aqueous systems  
251 listed above, IPA and hexane.

252

253 Similarly, the extractions performed in the PQRI OINDP study, including Soxhlet and reflux,  
254 were consistent with the nature of the test materials, the extraction solvents and the nature of  
255 OINDP products. Because a significant portion of PODP products are water-based, extractions  
256 performed in this study will be include the OINDP methods and extraction methods compatible  
257 with aqueous extraction media, specifically sealed vessel extraction.

258

259 The specific operational details associated with performing these extractions are outlined in the  
260 following sections. Note that the outlined extraction parameters and conditions maybe subject to  
261 modification and the details of any modified extraction process will be established in  
262 consultation with study coordinator prior to initiation of experimental work in any particular  
263 laboratory. Additionally, all extractions should be performed with appropriate extraction blanks.

264

265

266 **B. Extraction Maps**

267  
 268 The number of potential test situations, defined as the coupling of a test material, an extraction  
 269 solvent and an extraction process, is large and addressing each individual test situation is not  
 270 necessary to generate relevant information upon which best demonstrated practice  
 271 recommendations may be based. Additionally, some experience has already been gained during  
 272 the characterization of the initial set of five test Articles. Finally, the nature of the label itself is  
 273 such that it is clear that under certain extraction procedures, the label would dissolve. Thus not  
 274 all extraction conditions utilized in the initial phase of this Study will be used to characterize the  
 275 two new test materials. Test situations that are within the scope of this study are delineated in  
 276 the following Extraction Maps. The intent of this Stage 1 assessment is to generate the  
 277 fundamental information from which best demonstrated practices can be derived; it is not the  
 278 intent of this Stage 1 assessment to establish the practices used in this study as best demonstrated  
 279 practices themselves.

280  
 281 *1. Test Material Versus Extraction Solvent Map*

282  
 283 Table 2 establishes which extraction solvents will be utilized with which materials.

284  
 285

Table 2. Material Versus Extraction Solvent Map (1, 3)						
	Aqueous		Mixed	Organic		Thermal (2)
	pH 2.5	pH 9.5	IPA/Water	IPA	Hexane	
LDPE and PP	X	X	X <sup>4</sup>	X	X	X
Label	X	X	X	---	---	X

286  
 287 Notes: (1) An X denotes a material/solvent couple that will be performed, an --- denotes a couple that will not be  
 288 performed.  
 289 (2) By Headspace analysis.  
 290 (3) During the course of this study it may be the case that certain material – solvent couples will be  
 291 incompatible. Such incompatibilities should be reported the PODP study coordinator and incompatible  
 292 extracts should not be tested.  
 293 (4) Both reflux and sealed vessel with the IPA/Water mixture

294  
 295 *2. Extraction Method Versus Extraction Solvent Map*

296  
 297 Table 3 establishes which extraction methods will be utilized with which extraction solvents.

298  
 299  
 300  
 301  
 302  
 303  
 304  
 305



Table 3. Extraction Method Versus Extraction Solvent Map (1, 4)					
	Aqueous		Mixed IPA/Water	Organic	
	pH 2.5	pH 9.5		IPA	Hexane
<b>Testing for the LDPE and PP</b>					
Soxhlet	---	---	---	X	X
Reflux	---	---	--	X	X
Sealed Vessel	X (2)	X (2)	X (3)	---	--
<b>Testing for the Label</b>					
Sealed Vessel	X (2)	X (2)	X (3)	---	--

306  
 307 Notes: (1) An X denotes a method/solvent couple that will be performed, an --- denotes a couple that will not be  
 308 performed.  
 309 (2) Under autoclave conditions (121°C for 1 hr).  
 310 (3) Storage at 55°C for 3 days.  
 311 (4) During the course of this study it may be the case that certain material – solvent couples will be  
 312 incompatible. Such incompatibilities should be reported the PODP study coordinator and incompatible  
 313 extracts should not be tested.  
 314

315 Soxhlet and reflux extractions will not be performed on the label as it is envisioned that such  
 316 extractions would essentially dissolve the label. Sonication extractions will not be used in this  
 317 phase of the study as the previous study results suggest that this method does not produce useful  
 318 extractables profiles.  
 319

320 **C. General Considerations**

321  
 322 Care in experimental approach should be exercised in terms of producing extracts that are free  
 323 from analytical artifacts. Glass is the appropriate vessel for samples intended for organic  
 324 analysis, while Teflon is recommended for inorganic (metals) analysis. Glass is a problem in  
 325 metal analysis especially at higher pHs due to leaching of glass (e.g. Si, B, Al, Na). Teflon is a  
 326 problem with organics due to adsorption of extractables.  
 327

328 Extraction vessels shall be cooled and the materials separated from the liquid, by an appropriate  
 329 means. The extracts shall be collected and stored in an appropriate vessel with minimal  
 330 headspace. Retain the extract for analysis in such a way as to preserve their compositional  
 331 integrity (protect from light, heat and evaporation losses).  
 332

333 For all extractions, the weight of test article sample, extracting solvent volume, and sample  
 334 extract concentration factors should be established and adjusted so that it is possible to detect and  
 335 identify individual extractables present at the 10 µg/g (ppm) level. Individual extractables may  
 336 be detected and identified at lower levels if the analytical method employed is readily capable of  
 337 achieving such sensitivity.  
 338

339 For each extraction technique and solvent type, appropriate blanks (no test article sample) must  
 340 be prepared. These must be prepared concurrently using a different extraction apparatus (same  
 341 type) under the same conditions, or by using the same apparatus prior to charging with sample.  
 342 The extraction conditions represent the consensus opinion of the PODP chemistry subteam.

343  
344 All extracts should be visually inspected prior to analysis to ensure that they are free from  
345 obvious particulate matter. Should such an inspection reveal particulate matter, this finding  
346 should be reported to the Study Coordinator prior to proceeding with sample analysis. In most  
347 cases it is likely that the Study Coordinator will request that the sample be processed in such a  
348 way that the particulate is removed from the extract prior to its testing. Collection of the  
349 removed particulate may be requested so that the material itself can be analyzed and identified.  
350

#### 351 **D. Soxhlet Extraction**

352  
353 The conditions for performing Soxhlet extractions were specified in the original Protocol. Any  
354 modifications appropriate to the conditions specified in the original Protocol, based on the  
355 experiences gained during the characterization of the five original Test Articles, will be  
356 documented in the Final Report associated with this study.  
357

#### 358 **E. Reflux**

359  
360 The conditions for performing Reflux extractions were specified in the original Protocol. Any  
361 modifications appropriate to the conditions specified in the original Protocol, based on the  
362 experiences gained during the characterization of the five original Test Articles, will be  
363 documented in the Final Report associated with this study.  
364

#### 365 **F. Sealed Vessel Extraction**

366  
367 The conditions for performing sealed vessel extractions were specified in the original Protocol.  
368 Any modifications appropriate to the conditions specified in the original Protocol, based on the  
369 experiences gained during the characterization of the five original Test Articles, will be  
370 documented in the Final Report associated with this study.  
371

### 372 **VIII. ANALYTICAL METHODS**

#### 373 374 **A. General**

375  
376 Considerable experience was gained during the characterization of the original five Test Articles,  
377 specifically related to the analytical methods employed. While in general the same analytical  
378 techniques outlined in the original Protocol will be used with the additional two Test Articles, the  
379 specific operating details of the methods used may be somewhat different from those specified in  
380 the original Protocol. Any modifications appropriate to the conditions specified in the original  
381 Protocol, based on the experiences gained during the characterization of the five original Test  
382 Articles, will be documented in the Final Report associated with this study.  
383

384 The system suitability requirements contained in the original Protocol are relevant to and  
385 required for the analyses performed to characterize the two additional Test Articles.  
386  
387

388 **B. Gas Chromatography (GC)**  
389

390 The conditions for performing GC analyses of the extracts were specified in the original  
391 Protocol. Any modifications appropriate to the conditions specified in the original Protocol,  
392 based on the experiences gained during the characterization of the five original Test Articles,  
393 will be documented in the Final Report associated with this study.  
394

395 **C. High Performance Liquid Chromatography (HPLC)**  
396

397 The conditions for performing HPLC analyses of the extracts were specified in the original  
398 Protocol. Any modifications appropriate to the conditions specified in the original Protocol,  
399 based on the experiences gained during the characterization of the five original Test Articles,  
400 will be documented in the Final Report associated with this study.  
401

402 **D. Inductively Coupled Plasma – Mass Spectrometry (ICP-MS)**  
403

404 The conditions for performing ICP-MS analyses of the extracts were specified in the original  
405 Protocol. Any modifications appropriate to the conditions specified in the original Protocol,  
406 based on the experiences gained during the characterization of the five original Test Articles,  
407 will be documented in the Final Report associated with this study.  
408

409 **E. Headspace GC/MS**  
410

411 The conditions for performing Headspace GC/MS analyses of the Test Articles themselves were  
412 specified in the original Protocol. Any modifications appropriate to the conditions specified in  
413 the original Protocol, based on the experiences gained during the characterization of the five  
414 original Test Articles, will be documented in the Final Report associated with this study.  
415

416 **IX. DATA EVALUATION AND REPORTING**  
417

418 **A. Qualitative Analysis**  
419

- 420 • A list of all identified entities (compounds, elements) that were not detected in the  
421 corresponding blank. This list should include the recognized compound name,  
422 CAS Registry number, chemical formula, and chemical structure.
- 423 • A list of all unidentified chromatographic peaks that were not detected in the  
424 corresponding blank at signal to noise ratios greater than 10. The participating  
425 laboratory should determine and report the analyte concentration that corresponds  
426 to this signal to noise ratio (typically defined as the limit of quantitation, LOQ).
- 427 • Copies of chromatograms, spectra, etc.
- 428 • Complete methodological information for both the extraction and analysis  
429 processes.
- 430 • The required system suitability results, which should include an assessment of  
431 detectability.

- 432           • The identification status for all compounds shall be established and reported as  
433 follows:  
434
- 435           • A *Confirmed* identification means that collaborating information has been  
436 obtained including mass spectrometric fragmentation pattern, confirmation of  
437 molecular weight (or elemental composition), match in retention time and  
438 spectrum with authentic standard.
  - 439           • A *Confident* identification means that sufficient data to preclude all but the most  
440 closely related structures have been obtained
  - 441           • A *Tentative* identification means that data have been obtained that are consistent  
442 with a class of molecule only.  
443

#### 444 B. Semi-Quantitative Analysis

445  
446 While it is not the primary intent of this Stage 1 Protocol to produce quantitative data,  
447 some of the test methods employed may be amenable to concentration estimation (e.g.  
448 ICP, GC with internal standards). In the case that a participating laboratory reports  
449 concentration estimates, the means by which such estimates were obtained must be  
450 indicated. Additionally, all such estimates shall be reported with a convention (e.g.  
451 significant figures) which effectively reflects the uncertainty in the determination. As  
452 was noted previously, the threshold for reporting semi-quantitative results is 10 µg/g.  
453

#### 454 X. GLOSSARY OF ABBREVIATIONS

455	GC/FID	Gas Chromatography with Flame Ionization Detector
	GC/MS	Gas Chromatography with Mass Spectrometric Detection
	HPLC/DAD	High Pressure Liquid Chromatography-Diode Array Detection
	LC/MS	Liquid Chromatography Mass Spectrometric Detection
	ICP-MS	Inductively Coupled Plasma Atomic Emission Spectroscopy
	TIC	Total Ion Chromatogram
	API-ES	Atmospheric Pressure Ionization - Electrospray
	HS	Headspace
456	PQRI	Product Quality Research Institute
457	OINDP	Orally Inhaled and Nasal Drug Products
	PODP	Parenteral and Ophthalmic Drug Products
458	LDPE	Low density polyethylene
459	PP	Polypropylene
460	LVP	Large volume parenteral
461	SVP	Small volume parenteral
462	BFS	Blow-fill-seal
463	PFS	Pre-filled syringe
464		
465		

466 **XI. REFERENCES**

467  
468 PQRI Research Project Proposal: *Development of Scientifically Justifiable Thresholds and Best*  
469 *Demonstrated Characterization Practices for Leachables and Extractables in Parenterals and*  
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475 European Medicines Agency (EMA): *Guideline on Plastic Immediate Packaging Materials*,  
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479 *Experimental Protocol for Qualitative Controlled Extraction Studies on Material Test Articles*  
480 *Representative of Prefilled Syringe (PFS) and Small Volume Parenteral (SVP) Container*  
481 *Closure Systems*. Study Protocol – Stage 1. Issued and Effective, December, 2009.

482  
483